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Center for Veterinary Biologics**

SAM 635

**Supplemental Assay Method for Potency Testing of *Pasteurella multocida*
Bacterins of Porcine Origin**

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Contact: Janet M. Wilson, (515) 663-7245

Approvals: /s/Paul J. Hauer Date: 22Jan07
Paul J. Hauer, Section Leader
Bacteriology

/s/Byron E. Rippke Date: 30Jan07
Byron E. Rippke, Director
Policy, Evaluation, and Licensing
Center for Veterinary Biologics

/s/Rebecca L.W. Hyde Date: 31Jan07
Rebecca L.W. Hyde, Section Leader
Quality Assurance
Center for Veterinary Biologics

United States Department of Agriculture
Animal and Plant Health Inspection Service
P. O. Box 844
Ames, IA 50010

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Supplemental Assay Method for Potency Testing of *Pasteurella multocida* Bacterins of Porcine Origin

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1. Introduction

This Supplemental Assay Method (SAM) describes procedures for potency testing biological products containing *Pasteurella multocida* of porcine origin, as prescribed in the Code of Federal Regulations, Title 9 (9 CFR), Part 113.121. Mice are vaccinated twice, 14 days apart, and challenged with a standard dose of virulent *P. multocida* 10 to 12 days after the second vaccination.

2. Materials

2.1 Equipment/instrumentation

Equivalent equipment or instrumentation may be substituted for any brand name listed below.

2.1.1 Spectrophotometer, Spectronic 20D+ (Spectronic Instruments)

2.1.2 Sterile inoculating loop

2.1.3 Bunsen burner (if nonsterile wire inoculating loop is used)

2.1.4 Incubator, 35°- 37°C

2.1.5 Micropipettors, 20- to 1000-µL

2.1.6 Test tube mixer; vortex type

2.1.7 Crimper for aluminum caps on serum vials

2.1.8 Biological safety cabinet

2.2 Reagents/supplies

Equivalent reagents or supplies may be substituted for any brand name listed below.

2.2.1 *P. multocida* challenge culture, Strain 169, available from the Center for Veterinary Biologics (CVB). Refer to the current reagent data sheet for additional information.

2.2.2 Test bacterin(s) containing *P. multocida*

2.2.3 *P. multocida* reference bacterin, available from the CVB. Refer to the current reagent data sheet for additional information.

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- 2.2.4 Syringes, 1-mL
- 2.2.5 Needles, 26-gauge, 3/8-inch
- 2.2.6 Glass serum bottle, 20- to 100-mL
- 2.2.7 Rubber stopper, 13 x 20-mm, and aluminum cap for serum bottle
- 2.2.8 Glass screw-cap tubes, 13 x 100-mm
- 2.2.9 Pipettes, 5-mL, 10-mL, 25-mL
- 2.2.10 Micropipette tips, up to 1000- μ L capacity
- 2.2.11 Bovine blood agar plates
- 2.2.12 Tryptose broth
- 2.2.13 Phosphate-buffered saline (PBS)
- 2.2.14 Water, distilled or deionized, or water of equivalent purity

2.3 Animals

2.3.1 Mice, 16-22 g. Although the 9 CFR does not specify a specific mouse type, the CVB uses CF-1 mice.

2.3.2 Sixty mice are required for each bacterin to be tested (20 mice/dilution; 3 dilutions/bacterin). Sixty additional mice are required for the reference bacterin. Thirty mice are required to determine the LD₅₀ of the challenge inoculum. All mice must be from the same source colony and of similar weight and/or age.

3. Preparation for the test

3.1 Personnel qualifications/training

Technical personnel must have a working knowledge of the use of general laboratory chemicals, equipment, and glassware and must have specific training and experience in sterile technique, the handling of live bacterial cultures, and the handling of mice.

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3.2 Selection and handling of test animals

3.2.1 Mice of either sex may be used, but females are recommended.

3.2.2 All mice must be housed and fed in a similar manner.

3.2.3 Identify each cage of mice by treatment group.

3.2.4 If any mice die after vaccination, but prior to challenge with live *P. multocida*, perform a necropsy on these mice to determine cause of death if the cause of death is not outwardly apparent. If the cause of death is unrelated to vaccination, file the necropsy report with the test records, and no additional action is needed. If death is attributable to the test bacterin, report the death immediately to the CVB-Inspection and Compliance, which may request further safety testing of the bacterin.

3.2.5 When the test is concluded, instruct the animal caretakers to euthanize and incinerate the mice and to sanitize the contaminated rooms.

3.3 Preparation of supplies/equipment

3.3.1 Use only sterile supplies.

3.3.2 Operate and maintain all equipment according to manufacturers' recommendations and applicable standard operating procedures.

3.4 Preparation of reagents

3.4.1 *P. multocida* reference bacterin. Refer to the current reagent data sheet for details.

3.4.2 *P. multocida* challenge culture, Strain 169. Refer to the current reagent data sheet for details.

3.4.4 Phosphate-buffered saline--National Veterinary Services Laboratories (NVSL) Media #10559

Sodium chloride	8.0 g
Potassium chloride	0.2 g
Sodium phosphate, dibasic	1.15 g
Potassium phosphate, monobasic	0.2 g
Deionized water	q.s. to 1000 mL

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Adjust pH to 7.2 ± 0.1 . Autoclave 20 minutes at $\geq 121^\circ\text{C}$. Store at 20° - 25°C for no longer than 6 months.

3.4.5 Tryptose broth--NVSL Media #10404

Tryptose broth powder	26 g
Deionized water	q.s. to 1000 mL

Autoclave 15 minutes at $\geq 121^\circ\text{C}$. Cool before using. Store at 20° - 25°C for no longer than 6 months.

3.4.6 5% bovine blood agar--NVSL Media #10006

Blood agar base powder	40 g
Water	q.s. to 950 mL

Autoclave 20 minutes at $\geq 121^\circ\text{C}$. Cool to 45° - 47°C .

Add:

Defibrinated bovine blood	50 mL
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Pour into sterile petri dishes. Cool to 20° - 25°C . Store at 2° - 7°C for no longer than 6 months.

4. Performance of the test

4.1 Vaccination of test animals

4.1.1 Check the label on each product and Section VI of the current Outline of Production to confirm identity and dose volume.

4.1.2 Test each test bacterin and the reference bacterin at 3 fivefold dilutions. Typically, test the bacterins undiluted, 1:5, and 1:25. It is permissible to make fivefold dilutions other than those described as long as the reference and test bacterins are tested at the same dilutions. For viscous bacterins, it is advisable to start at 1:2 or 1:3 and make fivefold dilutions from this starting point to increase injectability of the product at the low dilution.

4.1.3 Thoroughly mix product by inverting end-to-end at least 10 times. Make the appropriate fivefold dilutions of the reference bacterin in PBS. Make identical fivefold dilutions of the test bacterin(s) in the diluent approved in the specific Outline of Production for that product. (Some oil-adjuvanted products

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require oil-based diluents.) Place each dilution in a separate sterile injection vial. Prepare dilutions immediately prior to use; do not store in diluted form.

4.1.4 Vaccinate separate groups of 20 mice with each of the test bacterin dilutions and reference bacterin dilutions. For reference bacterin groups, inject each mouse with 0.1 mL intraperitoneally. Inject test bacterins intraperitoneally at a dose volume that corresponds to 1/20 of the lowest dose recommended on the product label. This volume must not be <0.1 mL.

Note: It is permissible to vaccinate a few extra mice in each group to compensate for any potential deaths that may occur prior to challenge and are not related to vaccination. However, if extra mice are vaccinated, all surviving at the time of challenge must be challenged with live *P. multocida* and included in data calculations.

4.1.5 Revaccinate the mice in a similar manner 14 days after the first vaccination.

4.1.6 Retain 30 nonvaccinated mice to determine LD₅₀ of the challenge.

4.2 Preparation of challenge in a biological safety cabinet

4.2.1 Reconstitute a vial of challenge in 1 mL tryptose broth.

4.2.2 Inoculate 2 blood agar plates with a loopful of reconstituted culture and streak for isolation.

4.2.3 Incubate the inoculated blood agar plates at 35°- 37°C for 16 to 18 hours.

4.2.4 Use plates that have pure growth by visual inspection to prepare the challenge inoculum.

4.2.5 Scrape several bacterial colonies from the surface of the blood agar plates and suspend in tryptose broth in a 13 x 100-mm tube. Add bacterial growth until the suspension measures 76-80% T at 630 nm using a Spectronic 20D+ spectrophotometer or equivalent. Use sterile tryptose broth in a 13 x 100-mm tube as a blank for the spectrophotometer.

4.2.6 Prepare a 10⁻⁴ dilution of the standardized culture in tryptose broth. **This is the inoculum used to challenge the mice.** Dispense challenge liquid in a serum vial and seal with a rubber stopper and aluminum ring.

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4.2.7 Make additional tenfold dilutions for challenge LD₅₀ determination (10⁻⁶ to 10⁻⁸) and postinoculation plate counts (10⁻⁵ to 10⁻⁷). Dispense an aliquot of each LD₅₀ dilution in a separate vial and seal.

4.2.8 Place all vials of challenge inoculum and additional dilution tubes on ice. Keep on ice through the challenge procedure and until culture is added to plates for postinoculation plate count.

4.3 Timing and administration of challenge

4.3.1 Challenge all vaccinates 10 to 12 days after the second vaccination.

4.3.2 Challenge nonvaccinated LD₅₀ controls at the same time as the vaccinates.

4.3.3 Inoculate each vaccinated mouse with 0.2 mL of challenge inoculum intraperitoneally, using a 1-mL syringe and 26-gauge, 3/8-inch needle.

4.3.4 Inoculate separate groups of 10 nonvaccinated control mice intraperitoneally with 0.2 mL of each of the LD₅₀ dilutions.

4.4 Postinoculation plate count in a biological safety cabinet

4.4.1 All bacterial suspensions must be mixed well prior to placing an aliquot on an agar plate. Plate each dilution (10⁻⁵ to 10⁻⁷) in triplicate using 0.1 mL on bovine blood agar. Inoculum must be spread evenly on the surface of the agar plates and not allowed to pool around the edges. Complete all plate inoculations within 1 hour of challenge.

4.4.2 Incubate the plates aerobically at 35°- 37°C for 18 to 30 hours.

4.4.3 Using the dilution yielding 30-300 colonies per plate, calculate the colony forming units (CFU)/challenge dose according to the following formula:

$$\frac{\text{Colony count sum}}{\text{Number of plates}} \times \frac{1}{\text{Dilution factor plated}} \times \frac{1}{\text{Plated volume (mL)}} \times \frac{\text{Challenge dilution}}{1} \times \frac{\text{Challenge vol. (mL)}}{\text{Dose}} = \frac{\text{CFU}}{\text{Dose}}$$

4.4.4 Record the plate count (CFU/dose) of the challenge on the test result form. This information is for informational purposes to track trends and to troubleshoot problem tests. The 9 CFR does not specify a minimum or maximum CFU/dose for this test.

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4.5 Observation of mice after challenge

4.5.1 Observe the mice daily for 10 days after challenge. Record deaths.

4.5.2 If deaths occurring after challenge are suspected to be due to causes other than pasteurellosis, perform a necropsy on such mice to determine the cause of death. If cause of death is unrelated to vaccination and/or challenge, do not include the deaths in the total deaths for the test.

5. Interpretation of the test results

Interpret the test as prescribed in 9 CFR, Part 113.121.

5.1 Calculate the LD₅₀ (theoretical dose/dilution at which the challenge would be lethal to 50% of the control mice) of the challenge inoculum using the Reed-Muench or Spearman-Kärber method of estimation. A valid test must have an LD₅₀ between 100 and 10,000.

5.2 Calculate the PD₅₀ of the reference bacterin and each test bacterin (theoretical dose/dilution at which the bacterin would protect 50% of the mice) using the Reed-Muench or Spearman-Kärber method of estimation.

5.3 If the PD₅₀ of the reference bacterin cannot be calculated because the lowest dilution protects <50% of the mice or the highest dilution protects >50% of the mice, the test is invalid. The reference bacterin also must protect >0% and <100% of the mice at 2 or more dilutions in a valid test.

5.4 If the PD₅₀ of the test bacterin cannot be calculated because the lowest dilution tested protects <50% of the mice, the bacterin may be retested, *provided the following*:

- 1.** If the bacterin is not retested, it is unsatisfactory.
- 2.** If the protection provided by the lowest dilution of the reference exceeds that provided by the lowest dilution of the test bacterin by at least 6 mice, the test bacterin is unsatisfactory without additional testing.
- 3.** If the total number of mice protected by the reference (sum of survivors in all dilution groups) exceeds the total number protected by the test bacterin by 8 mice or more, the test bacterin is unsatisfactory without additional testing.

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5.5 If the PD₅₀ of the test bacterin in a valid test cannot be calculated because the highest dilution protected more than 50% of the mice, the test bacterin is satisfactory without further testing.

5.6 Divide the PD₅₀ of each test bacterin by the PD₅₀ of the reference to calculate the relative potency (RP) for each test bacterin.

5.7 If the RP of the test bacterin is ≥ 0.50 , the test bacterin is satisfactory.

5.8 If the RP of the test bacterin is < 0.50 , the test bacterin is unsatisfactory.

5.9 A test bacterin with an RP < 0.50 may be retested by conducting 2 independent replicate tests in a manner identical to the initial test. Calculate the results of the retests in the following manner:

- 1.** Average the RP values of the retests.
- 2.** If the average RP of the retests is < 0.50 , the bacterin is unsatisfactory.
- 3.** If the average RP of the retests is ≥ 0.50 **AND** the RP obtained in the original test is $\leq 1/3$ than the average (RP) of the retests, the test bacterin is satisfactory. Consider the initial test to be the result of test system error.
- 4.** If the average of the retests is ≥ 0.50 **BUT** the RP of the original test is $> 1/3$ of the average RP of the retests, calculate a new average RP using the RP values obtained in all tests (original plus retests). If the new average RP is ≥ 0.50 , the test bacterin is satisfactory. If the new average RP is < 0.50 , the test bacterin is unsatisfactory.

6. Report of test results

Report results of the test(s) as described by standard operating procedures.

7. References

7.1 Code of Federal Regulations, Title 9, Part 113.121, U.S. Government Printing Office, Washington, DC, 2006.

7.2 Reed LJ, Muench H, 1938. A simple method of estimating 50% endpoints. *Am J Hygiene*, 27:493-497.

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7.3 Cottral G.E., (Ed.), 1978. *Manual of standardized methods for veterinary microbiology*. Comstock Publishing Associates, Ithaca, NY. pg. 731.

7.4 Finney, D.J. 1978. *Statistical methods in biological assay*. Griffin, London. 3rd edition, pp. 394-401.

8. Summary of revisions

Version .04

- **4.2.7** The LD₅₀ dilutions have been adjusted to reflect current practices.
- **7.3** and **7.4** Additional references have been added.

Version .03

- **2.1** A sterile inoculating loop and biological safety cabinet have been added.
- **4.2.2** The number of plates used has been updated to reflect current practices.
- **4.2.6** The current dilution has been updated to reflect current practices.
- References to the current reagent data sheet have been added throughout the document.
- References to internal documents have been replaced with summarized information.
- Parameters for the currently used spectrophotometer have been updated.
- The contact person has been changed to Janet Wilson.